

Indole Alkaloids Produced by a Marine Fungus Isolate of *Penicillium janthinellum* Biourge

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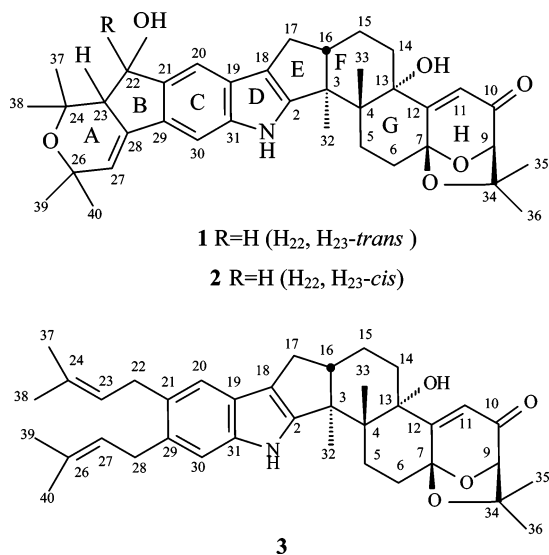
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Three new indole alkaloids, shearinines D, E, and F (**1–3**), together with the known shearinine A (**4**) were isolated from the marine-derived strain of the fungus *Penicillium janthinellum* Biourge. The chemical structures of **1–4** were established by 2D NMR and HREIMS data. Shearinines A, D, and E induce apoptosis in human leukemia HL-60 cells, and shearinine E also inhibits EGF-induced malignant transformation of JB6 P⁺ Cl 41 cells in a soft agar.

Marine-derived fungi are a rich source of structurally new natural products with a wide range of biological activities.^{1–3} In our search for new compounds from marine-derived fungi, three new indole-diterpenoid alkaloids, shearinines D, E, and F (**1–3**), together with the known shearinine A (**4**)⁴ were obtained from the fungus *Penicillium janthinellum* Biourge, which was isolated from marine sediments. These metabolites are analogues of paspalinine (**5**)^{5,6} and janthitrems B–G (**6a**, **6b**, **7a**, **7b**, **7c**)^{7,8} previously discovered from terrestrial fungi. Here we describe the isolation and structural elucidation of new alkaloids produced by the marine fungus *P. janthinellum*.

Results and Discussion

The EtOAc extract of the culture of *P. janthinellum* was fractionated by Si gel column chromatography using a step gradient–elution technique employing mixtures of hexane–EtOAc as solvent to yield individual compounds **1–4**. The structures of compounds **1–4** were established by the interpretation of physical data (¹H NMR, ¹³C NMR, HSQC, HMBC, COSY, NOESY, and HREIMS). The physical data of the compounds were compared with literature data of the related compounds.^{4–6}



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The molecular formula of compound **1** was determined as C₃₇H₄₅O₆N on the basis of HREIMS and ¹³C NMR analysis. Inspection of the ¹H and ¹³C NMR data (Table 1) of **1** revealed the presence of eight quaternary methyls (δ 16.2, C-32; 23.6, C-33; 23.1, C-35; 28.8, C-36; 30.0, C-37; 23.0, C-38; 31.9, C-39; 30.0, C-40), five methylenes (δ 27.0, C-5; 28.2, C-6; 33.9, C-14; 21.1, C-15; 27.5, C-17), six methines (δ 88.0, C-9; 117.7, C-11; 48.5, C-16; 76.4, C-22; 60.1, C-23; 120.9, C-27) including two olefinic and two oxygen-bearing ones, two aromatic proton singlets (δ _H 7.44, H-20; 7.34, H-30), and an N-H singlet (δ _H 7.71). Absorptions at 233 nm (log ϵ 4.1), 256 (sh) (4.4), 263 (4.3), and 330 (4.1) in the UV spectrum of the shearinines, e.g., shearinine D (**1**), suggested the presence of a 2,3-disubstituted indole nucleus with a double bond in conjugation with the aromatic ring system as in janthitrems E, F, and G⁸ and shearinine A.⁴ The general features of the ¹H and ¹³C NMR spectra (Table 1) of **1** closely resembled those of shearinine A (**4**),⁴ with the exception of proton and carbon resonances corresponding to one additional secondary hydroxy group (δ _H 4.96, δ _C 76.4). HMBC correlations between H-22 and C-20, C-28, and C-29 placed this hydroxy group at C-22 of ring B. The NOESY spectrum exhibited cross-peaks H-22/H₃-38, indicating that these protons are cofacial (Figure 1). The cross-peak between H₃-38 and H-23 in the COSY-45 spectrum demonstrated that these protons were oriented on opposite faces of the molecule. The coupling constants⁹ of the resonances at δ 4.96 (H-22) and 2.67 (H-23) indicated that the gross structure of shearinine D (**1**) was H₂₂,H₂₃-*trans*-22-hydroxyshearinine A.

The relative configuration of **1** was clarified by analysis of the NOESY spectrum. Thus, from the presence of NOESY correlations from H-16 to H₃-33 and from H₃-32 to H-17 α and the absence of the NOE between H-16 and H₃-32 the *trans*-3,16-ring junction was deduced. The steric relationship between the indole portion of **1** and the diterpene part is determined by NOESY correlations between H-20 and H-17.

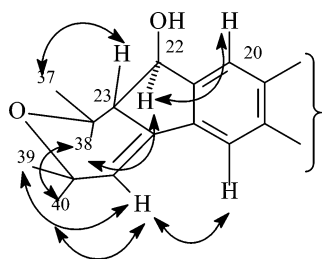
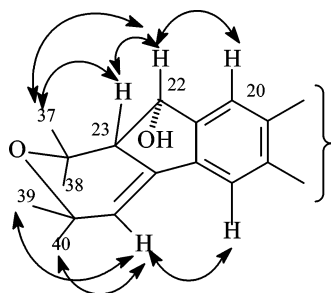
Shearinine E (**2**) has the same molecular formula as shearinine D (**1**). The NMR data of **2** were very similar to those obtained for shearinine D (Tables 1, 2) with the exception of the C-22 and C-23 resonances (Tables 1, 2). The NOESY spectrum revealed correlations between H-22 and H-23, H-20, and H₃-37. The coupling constants¹⁰ of the resonances at δ 4.96 (H-22) and 2.67 (H-23) (Figure 2) suggested that **2** is an isomer of **1**. Therefore, the structure of shearinine E (**2**) was determined as H₂₂,H₂₃-*cis*-22-hydroxy-shearinine A.

The molecular formula of compound **3** was determined as C₃₇H₄₇O₄N on the basis of HREIMS and NMR spectra. The general features of the ¹H and ¹³C NMR spectra of **3** (Table 3) closely resembled those of shearinines D (**1**) and E (**2**) (Tables 1, 2) and also of 21-isopentenylpaxilline⁴ and paspalinine (**5**),⁵ with the

Table 1. NMR Spectroscopic Data (500 MHz, CDCl₃) for Shearinine D (1)

| position | δ_C | DEPT | δ_H (J in Hz) | NOESY (350 ms) and 1D NOE | HMBC ^a |
|----------|--------------------|-----------------|---|---------------------------|-------------------------------|
| 1(NH) | | | 7.71 brs | 5, 33, 30 | 2, 18, 19, 31 |
| 2 | 153.8 | C | | | |
| 3 | 51.7 | C | | | |
| 4 | 39.9 | C | | | |
| 5 | 27.0 | CH ₂ | 2.69 m 1.82 m | 32 1 | 3, 4, 6, 7 3, 4, 6 |
| 6 | 28.2 | CH ₂ | 2.81 m 2.05 m | | 7, 5 7, 12, 4 |
| 7 | 104.3 | C | | | |
| 9 | 88.0 | CH | 4.32 d (1.3) | 35, 36 | 7, 10, 11, 36 |
| 10 | 196.9 | C | | | |
| 11 | 117.7 | CH | 5.84 brs | 14 | 7, 9, 10, 13 |
| 12 | 169.5 | C | | | |
| 13 | 77.6 | C | | | |
| 14 | 33.9 | CH ₂ | 2.00 m | 11 | |
| 15 | 21.1 | CH ₂ | 2.06 m 1.80 m | 32 | |
| 16 | 48.5 | CH | 2.82 m | 33 | |
| 17 | 27.5 | CH ₂ | 2.75 dd (13.2, 6.2) 2.44 dd (13.1, 10.5) | 32 | 2, 3, 16, 18 2, 15, 16, 18 |
| 18 | 117.5 | C | | | |
| 19 | 126.9 | C | | | |
| 20 | 113.9 | CH | 7.44 s | 22, 17 | 18, 19, 29, 30, 31 |
| 21 | 138.7 ^b | C | | | |
| 22 | 76.4 | CH | 4.96 brd (5.3) | 20, 37, 38 | 20, 24, 28, 29 |
| 23 | 60.1 | CH | 2.67 dd (5.7, 2.9) | 37 | 22, 24, 27, 28, 37 |
| 24 | 73.8 | C | | | |
| 26 | 72.4 | C | | | |
| 27 | 120.9 | CH | 5.96 d (2.9) | 30, 39, 40 | 23, 26, 29, 39, 40 |
| 28 | 135.1 ^b | C | | | |
| 29 | 131.9 | C | | | |
| 30 | 102.6 | CH | 7.34 s | 27, 1 | 19, 20, 21, 28 |
| 31 | 141.1 | C | | | |
| 32 | 16.2 | CH ₃ | 1.38 s | 5, 17a, 15 | 2, 3, 4, 16 |
| 33 | 23.6 | CH ₃ | 1.23 s | 1, 16, 14 | 3, 4, 5, 13 |
| 34 | 78.8 | C | | | |
| 35 | 23.1 | CH ₃ | 1.18 s | 9 | 9, 34, 36 |
| 36 | 28.8 | CH ₃ | 1.44 s | 9 | 9, 34, 35 |
| 37 | 30.0 | CH ₃ | 1.49 s | 38, 22, 23 | 23, 24, 38 |
| 38 | 23.0 | CH ₃ | 1.12 s | 40, 37, 22 | 23, 24, 37 |
| 39 | 31.9 | CH ₃ | 1.34 s | 27 | 26, 27, 40 |
| 40 | 30.0 | CH ₃ | 1.35 s | 38, 27 | 26, 27, 39 |

^a HMBC correlations, optimized for 8 Hz, are from proton(s) to the indicated carbon. ^b Assignments may be reversed.

**Figure 1.** Selected NOESY correlations in 1.**Figure 2.** Selected NOESY correlations in 2.

exception of proton and carbon resonances belonging to rings A and B. The resonances of four methyl groups at double bonds (δ_C 17.9– δ_H 1.74; δ_C 17.8– δ_H 1.72; δ_C 25.8– δ_H 1.75; δ_C 25.8– δ_H 1.74 ppm) and two methylene groups (δ_C 32.0– δ_H 3.39; δ_C 31.9–

Table 2. NMR Spectroscopic Data (500 MHz, CDCl₃) for Shearinine E (2)

| position | δ_C | DEPT | δ_H (J in Hz) | NOESY (350 ms) | HMBC ^a |
|----------|--------------------|-----------------|---|----------------|--------------------------------|
| 1(NH) | | | 7.72 brs | 33, 5, 30 | 2, 18, 19, 31 |
| 2 | 153.9 | C | | | |
| 3 | 51.7 | C | | | |
| 4 | 39.8 | CH ₂ | | | |
| 5 | 27.0 | CH ₂ | 2.70 brt (10.3) 1.83 m | 32 1, 33 | 3, 4, 6, 7, 33 3, 6, 13, 33 |
| 6 | 28.2 | CH ₂ | 2.81 m 2.05 m | | |
| 7 | 104.3 | C | | | |
| 9 | 87.9 | CH | 4.32 d (1.3) | 35, 36 | 7, 10, 11, 36 |
| 10 | 197.1 | C | | | |
| 11 | 117.7 | CH | 5.85 d (1.2) | 14 | 7, 9, 13 |
| 12 | 169.5 | C | | | |
| 13 | 77.6 | C | | | |
| 14 | 33.9 | CH ₂ | 2.00 td (12.4, 4.3) 1.90 m | 11, 33 | 13, 15, 16 |
| 15 | 21.1 | CH ₂ | 2.08 m 1.80 m | 32 | |
| 16 | 48.5 | CH | 2.82 m | 33 | |
| 17 | 27.5 | CH ₂ | 2.73 dd (6.3, 13.2) 2.44 dd (10.7, 13.2) | 20 32 | 2, 3, 1, 18 2, 15, 16, 18 |
| 18 | 117.6 | C | | | |
| 19 | 126.8 | C | | | |
| 20 | 114.9 | CH | 7.51 s | 22, 17 | 18, 22, 29, 30, 31, 19 |
| 21 | 139.9 ^b | C | | | |
| 22 | 74.3 | CH | 5.35 brd (7.0) | 20, 37, 23 | 20, 21, 28, 29 |
| 23 | 52.5 | CH | 2.90 dd (2.9, 7.3) | 22, 37 | 21, 24, 27, 37, 38 |
| 24 | 74.8 | C | | | |
| 26 | 72.8 | C | | | |
| 27 | 121.1 | CH | 6.04 d (2.9) | 39, 40, 30 | 23, 26, 29, 39, 40 |
| 28 | 136.7 ^b | C | | | |
| 29 | 132.5 | C | | | |
| 30 | 102.9 | CH | 7.38 s | 1, 27 | 19, 20, 21, 28 |
| 31 | 141.4 | C | | | |
| 32 | 16.0 | CH ₃ | 1.39 s | 5, 17, 15 | 2, 3, 4, 16 |
| 33 | 23.6 | CH ₃ | 1.23 s | 1, 16 | 3, 4, 5, 13 |
| 34 | 78.7 | C | | | |
| 35 | 23.1 | CH ₃ | 1.19 s | 9 | 9, 34, 36 |
| 36 | 28.9 | CH ₃ | 1.45 s | 9 | 9, 34, 35 |
| 37 | 29.8 | CH ₃ | 1.53 s | 22, 23 | 23, 24, 38 |
| 38 | 24.9 | CH ₃ | 1.38 s | | 23, 24, 37 |
| 39 | 32.2 | CH ₃ | 1.32 s | 27 | 26, 27, 40 |
| 40 | 29.3 | CH ₃ | 1.41 s | 27 | 26, 27, 39 |

^a HMBC correlations, optimized for 8 Hz, are from proton(s) to the indicated carbon. ^b Assignments may be reversed.

δ_H 3.39 ppm) in the ¹H and ¹³C NMR spectra and HMBC correlations of H-23 to C-37 and C-38 and H-27 to C-39 and C-40 proposed isopentenyl fragments connected at C-21 and C-29 atoms in 3. This supposition was confirmed by HMBC correlations of H-20 to C-18, C-22, C-29, and C-31 and H-30 to C-19, C-21, and C-28. Therefore, the structure of shearinine F (3) was determined as 21,22-diisopentenylpaspalinine. The relative configuration of ring fusions in 3 was the same as in 1 and 2 (based on the NOESY correlations: H-16/H-33, H-17/H-32, H-11/H-14, H-5 α /H-32).

The molecular formula of compound 4 was determined as C₃₇H₄₅NO₅ on the basis of HREIMS and NMR spectra. The remaining ¹H and ¹³C NMR assignments were similar to those of shearinine A.⁴ The connectivity of 4 and all ¹H and ¹³C NMR assignments were verified by analysis of HMBC, HSQC, and COSY experiments. The relative configuration of 4 was proposed on the basis of coupling constants, NOESY data, and structural identity to shearinine A.⁴ The good agreement of NMR spectra of 1 with those of 2, 3, and 4, together with NOESY data and proton coupling constants, supported that these compounds have identical relative configurations of ring fusions. The α -orientation of the hydroxy group at C-13 was supported by NOESY correlations, which were observed between H-17 α and H₃-32, and also between H₃-33 and H-16, and, additionally, NOE correlation between H-11 and H-14 α .

Shearinines A, D, E, and F were tested for cytotoxic properties against mouse epidermal JB6 P⁺ Cl 41 cells using the MTS method.¹¹ The compounds did not show any cytotoxicity up to 200 μ M concentration. Some of these compounds showed cancer preventive and antileukemic properties. Shearinine E inhibited EGF-

Table 3. NMR Spectroscopic Data (500 MHz, CDCl₃) for Shearinine F (3)

| position | δ_C | DEPT | δ_H (J in Hz) | NOESY (350 ms) | HMBC ^a |
|----------|--------------------|-----------------|--|----------------|---------------------------|
| 1(NH) | | | 7.52 brs | 33, 5, 30 | 2, 18, 19, 31 |
| 2 | 151.4 | C | | | |
| 3 | 51.4 | C | | | |
| 4 | 39.9 | C | | | |
| 5 | 26.9 | CH ₂ | 2.65 brt (10.5) 1.79 m | 32 1 | 6, 7, 33 |
| 6 | 28.2 | CH ₂ | 2.78 m 2.02 m | 33 | 5, 7 4, 7, 12 |
| 7 | 104.4 | C | | | |
| 9 | 88.0 | CH | 4.30 d (1.2) | 35, 36 | 7, 11, 36 |
| 10 | 197.1 | C | | | |
| 11 | 116.8 | CH | 5.83 brs | 14 | 7, 9, 13 |
| 12 | 169.8 | C | | | |
| 13 | 77.7 | C | | | |
| 14 | 33.9 | CH ₂ | 1.99 m 1.89 m | 11 11 | |
| 15 | 21.1 | CH ₂ | 2.05 m 1.77 m | 32 33 | |
| 16 | 48.5 | CH | 2.79 m | 33 | |
| 17 | 27.6 | CH ₂ | 2.70 dd (13.1, 6.3) (β) 2.40 dd (13.1, 10.6) (α) | 32 | 2, 3, 16, 18 2, 16, 18 |
| 18 | 116.8 | C | | | |
| 19 | 123.6 | C | | | |
| 20 | 118.4 | CH | 7.20 s | 22, 17 | 18, 22, 29, 31 |
| 21 | 131.3 | C | | | |
| 22 | 32.0 ^b | CH ₂ | 3.39 brd (7.1) | 20, 37 | 23, 24 |
| 23 | 124.3 ^b | CH | 5.31 m ^b | 38 | 37, 38 |
| 24 | 131.9 ^b | C | | | |
| 26 | 131.5 ^b | C | | | |
| 27 | 123.8 ^b | CH | 5.30 m ^b | 39 | 39, 40 |
| 28 | 31.9 ^b | CH ₂ | 3.39 brd (7.1) | 30, 40 | 26, 27 |
| 29 | 132.6 | C | | | |
| 30 | 111.4 | CH | 7.09 s | 1, 28 | 19, 21, 28 |
| 31 | 139.0 | C | | | |
| 32 | 16.2 | CH ₃ | 1.34 s | 5, 17, 15 | 2, 3, 4, 16 |
| 33 | 23.5 | CH ₃ | 1.21 s | 1, 16, 6 | 3, 4, 5, 13 |
| 34 | 78.7 | C | | | |
| 35 | 23.1 | CH ₃ | 1.18 s | 9 | 9, 34, 36 |
| 36 | 28.8 | CH ₃ | 1.44 s | 9 | 9, 34, 35 |
| 37 | 17.9 ^b | CH ₃ | 1.74 brs ^c | 22 | 23, 24, 38 |
| 38 | 25.8 | CH ₃ | 1.75 brs ^b | 23 | 23, 24, 37 |
| 39 | 25.8 | CH ₃ | 1.74 brs ^b | | 26, 27, 40 |
| 40 | 17.8 ^b | CH ₃ | 1.72 brs ^c | 28 | 26, 27, 39 |

^a HMBC correlations, optimized for 8 Hz, are from proton(s) to the indicated carbon. ^b · ^c Assignments may be reversed.

induced malignant transformation of JB6 P⁺ Cl 41 cells in a soft agar with INCC₅₀ (inhibition of number of the colonies₅₀) equal to 13 μ M concentration. The anchorage-independent transformation assay in a soft agar is a well-accepted tool to determine whether a compound can be a potentially effective cancer preventive agent in either humans or animals.^{12,13} Shearinines A, D, and E induced apoptosis in human leukemia HL-60 cells at 100 μ M concentration by 10%, 39%, and 34% of the apoptotic cells compared to control cells, respectively.

Experimental Section

General Experimental Procedures. Optical rotations were measured on a Perkin-Elmer 141 polarimeter, and melting points were taken with a Leica Galen III instrument. UV spectra were recorded on a Specord UV-vis spectrometer in MeOH. IR spectra were determined on a Specord M 82 Carl Zeiss Jena in CHCl₃. ¹H and ¹³C NMR spectra were recorded in CDCl₃ on a Bruker DRX-500 spectrometer operating at 500 and 125.6 MHz, respectively, using TMS as an internal standard. HREIMS were obtained with a JEOL MS Route mass spectrometer. MALDI MS analyses were carried out with a Bruker Biflex TOF mass spectrometer equipped with a UV-nitrogen laser (337 nm). Column chromatography was carried out using silica gel L (40/100 μ m, Chemapol, Czechoslovakia). Sephadex LH-20 was purchased from Pharmacia Biotech, Sweden. Plates (4.5 \times 6.0 cm) precoated with Si gel (5–17 μ m, Sorbfil, Russia) were used for thin-layer chromatography.

Cultivation of *P. janthinellum*. The fungus *P. janthinellum* was isolated from marine bottom sediments collected at a depth of 11 m (Sea of Japan, Amursky Bay, near Vladivostok). The fungus was

cultivated for 2 weeks at 30 °C in 20 0.5 L flasks, each containing 60 g of the nutrient medium RM¹⁴ of the following composition (g): sodium tartrate, 0.01; yeast extract, 0.02; rice, 20; KH₂PO₄, 0.01; seawater, 40.

Extraction and Isolation. The fungus mycelia with the medium were extracted for 24 h with 3 L of EtOAc. Evaporation of the solvent under reduced pressure gave a brown oil (3 g), to which 500 mL of H₂O–EtOH (4:1) was added, and the combination was thoroughly mixed to yield a suspension. It was extracted successively with hexane (500 mL \times 2), CHCl₃ (500 mL \times 2), and *n*-BuOH (500 mL \times 2). The CHCl₃ fraction was concentrated *in vacuo* to give a residue (700 mg), which was separated on a silica gel column (30 \times 3 cm) eluted with a hexane–EtOAc acetate gradient (hexane–EtOAc = 100:0; 95:5; 90:10; 87:13; 85:15; 80:20, each 1.5 L) to afford 10 fractions. Gel filtration of fractions 2, 4, 5, and 7 over a Sephadex LH-20 column with CHCl₃ gave **1** (10 mg), **2** (16 mg), **3** (8 mg), and **4** (60 mg).

Shearinine D (1): white solid (hexane–EtOAc), mp 180–185 °C (dec); [α]_D²⁰ +21.8 (*c* 0.13, CHCl₃); UV (CHCl₃) λ_{max} (log ϵ) 233 (4.13), 256 (sh) (4.36), 263 (4.34), 330 nm (4.08); IR (CHCl₃) ν_{max} 3585, 3471, 3013, 2979, 2933, 2865, 1688, 1602, 1456, 1377, 1365, 1354 cm⁻¹; HREIMS *m/z* 599.3273 (calcd for C₃₇H₄₅O₆N, 599.3247).

Shearinine E (2): white solid (hexane–EtOAc), mp >300 °C (dec); [α]_D²⁰ +3.5 (*c* 0.17, CHCl₃); UV (CHCl₃) λ_{max} (log ϵ) 231 (4.20), 256 (sh) (4.36), 264 (4.38), 286 (4.03), 330 nm (4.04); IR (CHCl₃) ν_{max} 3585, 3471, 3014, 2980, 2933, 2870, 1688, 1602, 1456, 1376, 1356 cm⁻¹; HREIMS *m/z* 599.3221 (calcd for C₃₇H₄₅O₆N, 599.3247).

Shearinine F (3): white solid (hexane–EtOAc), mp 128–130 °C; [α]_D²⁰ +76 (*c* 0.25, CHCl₃); UV (CHCl₃) λ_{max} (log ϵ) 234 (3.90), 300 nm (3.28); IR (CHCl₃) ν_{max} 3475, 3052, 3042, 3036, 3006, 2930, 2856, 1687, 1602, 1456, 1376, 1336 cm⁻¹; HREIMS *m/z* 569.3500 (calcd for C₃₇H₄₇O₄N, 569.3505).

Shearinine A (4): white solid (hexane–EtOAc), mp >300 °C (dec); IR (CHCl₃) ν_{max} 3582, 3474, 2979, 2937, 2871, 1688, 1455, 1377, 1365 cm⁻¹; MALDI-MS *m/z* 584 [M + H]⁺; other properties of **4** have been previously reported.⁴

Anchorage-Independent Transformation Assay. The mouse epidermal JB6 P⁺ Cl 41 cells were kindly donated by Prof. Zigang Dong (The Hormel Institute, University of Minnesota). The cancer preventive effect of shearinine E was evaluated using the JB6 P⁺ Cl 41 cell line and an anchorage-independent neoplastic transformation assay in a soft agar. EGF (10 ng/mL) was used for stimulating neoplastic transformation of JB6 P⁺ Cl 41 cells. The assay was carried out in six-well tissue culture plates. Mouse JB6 P⁺ Cl 41 cells (8 \times 10³/mL) were treated with various concentrations of shearinine E in 1 mL of 0.33% BME (basal medium Eagle) agar containing 10% FBS over 3 mL of 0.5% BME agar containing 10% FBS and various concentrations of shearinine B. The cultures were maintained in a 37 °C, 5% CO₂ incubator for 1 week. Cell colonies were then scored using a LEICA DM IRB inverted research microscope (Leica Mikroskopie und Systeme GmbH, Germany) and Image-Pro Plus software, version 3.0 for Windows (Media Cybernetics, Silver Spring, MD).

Apoptosis Assay Assessed by Flow Cytometry. The induction of early and late apoptosis by shearinines A, D, or E was analyzed by flow cytometry using a Becton Dickinson FACs caliber flow cytometer (BD Biosciences, San Jose, CA) after annexin V–fluorescein-isothiocyanate (FITC) and propidium iodide staining according to the manufacturer's protocol (BD Pharmingen, Franklin Lakes, NJ). In brief, HL-60 cells (2 \times 10⁵ cells/well) were grown in six-well plates for 12 h in 10% FBS/RPMI 1640 medium and then treated with 100 μ M concentration of shearinine A, D, or E for 24 h. After treatment with shearinines, the cells were washed and collected by centrifugation at 1000 rpm (170 rcf) for 5 min and resuspended in 500 μ L of 1 \times binding buffer. Then, 5 μ L of annexin V–FITC and 500 ng/mL of propidium iodide were added, and the cells were incubated at room temperature for 15 min in the dark and analyzed by flow cytometry.

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06-III-B-05, and 06-III-A-06-184, and by the Korea Science and Engineering Foundation (KOSEF) through the Medical Research Center for Cancer Molecular Therapy at Dong-A University (R13-2002-044-02003-0).

Supporting Information Available: Structural formulas of compounds 4–7. This material is available free of charge via the Internet at <http://pubs.acs.org>.

References and Notes

- (1) Blunt, J. W.; Copp, B. R.; Munro, M. H. G.; Northcote, P. T.; Prinsep, M. R. *Nat. Prod. Rep.* **2006**, *23*, 26–78.
- (2) Pivkin, M. V.; Kuznetsova, T. A.; Sova, V. V. *Marine Fungi and their Metabolites*; Dalnauka: Vladivostok, 2006; p 133.
- (3) Somei, M.; Yamada, F. *Nat. Prod. Rep.* **2005**, *22*, 73–103.
- (4) Belofsky, G. N.; Gloer, J. B.; Wicklow, D. T.; Down, P. F. *Tetrahedron* **1995**, *51*, 3959–3968.
- (5) Gallagher, R. T.; Finer, J.; Clardy, J.; Leutwiler, A.; Weibel, F.; Acklin, W.; Arigoni, D. *Tetrahedron Lett.* **1980**, *21*, 235–238.
- (6) Staub, G. M.; Gloer, K. B.; Gloer, J. B.; Wicklow, D. T.; Down, P. F. *Tetrahedron Lett.* **1993**, *34*, 2569–2572.
- (7) Penn, J.; Swift, R.; Wigley, L. J.; Mantle, P. G.; Bilton, J. N.; Sheppard, R. N. *Phytochemistry* **1993**, *32*, 1431–1434.
- (8) De Jesus, A. E.; Steyn, P. S.; Heerden, F. R.; Vleggaar, R. *J. Chem. Soc., Perkin Trans. 1* **1984**, 697–701.
- (9) Minale, L.; Pizza, C.; Zollo, F.; Riccio, R. *Tetrahedron Lett.* **1982**, *23*, 1841–1844.
- (10) Iorizzi, M.; Minale, L.; Riccio, R.; Debray, M.; Menou, J. L. *J. Nat. Prod.* **1989**, *52*, 1022–1024.
- (11) Barltrop, J. A.; Owen, T. C.; Cory, A. H.; Cory, J. G. *Bioorg. Med. Chem. Lett.* **1991**, *1*, 611–614.
- (12) Colburn, N. H.; Former, B. F.; Nelson, K. A.; Yuspa, S. H. *Nature* **1979**, *281*, 589–591.
- (13) Colburn, N. H.; Wendel, E. J.; Abruzzo, G. *Proc. Natl. Acad. Sci. U.S.A.* **1981**, *78*, 6912–6916.
- (14) Monaghan, R. L.; Polishook, J. D.; Pecor, V. J.; Bills, G. F.; Nallin-Omstead, M. *Can. J. Bot.* **1995**, *73*, 925–931.

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